

学校编码: 10384

密级

学号: 24520111153324

厦 门 大 学

硕 士 学 位 论 文

**microRNA 在皮肤和角膜上皮干细胞的差异性表达及其功能的初步研究**

**Differential expression of microRNAs between LSCs and  
SESCs and pilot study of their functions**

杜慧怡

指导教师姓名: 刘祖国 教授

李 程 助理教授

专 业 名 称: 眼科学

论文提交日期: 2014 年 4 月

论文答辩日期: 2014 年 5 月

2014 年 4 月

microRNA在角膜和皮肤干细胞的差异性表达及其功能的初步研究

杜慧怡

指导教师

刘祖国教授

李程助理教授

厦门大学

## 厦门大学学位论文原创性声明

本人呈交的学位论文是本人在导师指导下,独立完成的研究成果。本人在论文写作中参考其他个人或集体已经发表的研究成果,均在文中以适当方式明确标明,并符合法律规范和《厦门大学研究生学术活动规范(试行)》。

另外,该学位论文为( )课题(组)的研究成果,获得( )课题(组)经费或实验室的资助,在厦门大学眼科研究所及福建省眼科与视觉科学重点实验室完成。

声明人(签名):

年 月 日



## 厦门大学学位论文著作权使用声明

本人同意厦门大学根据《中华人民共和国学位条例暂行实施办法》等规定保留和使用此学位论文，并向主管部门或其指定机构送交学位论文（包括纸质版和电子版），允许学位论文进入厦门大学图书馆及其数据库被查阅、借阅。本人同意厦门大学将学位论文加入全国博士、硕士学位论文共建单位数据库进行检索，将学位论文的标题和摘要汇编出版，采用影印、缩印或者其它方式合理复制学位论文。

本学位论文属于：

（ ☒ ） 1.经厦门大学保密委员会审查核定的保密学位论文，  
于     年     月     日解密，解密后适用上述授权。

（        ） 2.不保密，适用上述授权。

（请在以上相应括号内打“√”或填上相应内容。保密学位论文应是已经厦门大学保密委员会审定过的学位论文，未经厦门大学保密委员会审定的学位论文均为公开学位论文。此声明栏不填写的，默认为公开学位论文，均适用上述授权。）

声明人（签名）



# 目录

|                                    |     |
|------------------------------------|-----|
| 英文缩略语对照表 .....                     | V   |
| 摘 要.....                           | VII |
| 第一章 前言 .....                       | 1   |
| 1.1 microRNA 的背景介绍.....            | 1   |
| 1.1.1 microRNA 的发现和功能介绍.....       | 1   |
| 1.2 角膜的正常解剖和生理 .....               | 3   |
| 1.3 角膜上皮细胞和皮肤上皮细胞 .....            | 4   |
| 1.3.1 角膜上皮和皮肤上皮的相似性和差异性.....       | 4   |
| 1.3.2 角膜上皮细胞和皮肤上皮细胞之间的相互转化.....    | 5   |
| 1.4 角膜上皮干细胞和皮肤上皮干细胞 .....          | 6   |
| 1.4.1 角膜上皮干细胞.....                 | 6   |
| 1.4.2 皮肤上皮干细胞.....                 | 8   |
| 1.5 microRNA 在角膜和皮肤上皮中的研究进展.....   | 9   |
| 1.5.1 microRNA 在角膜和皮肤上皮中重要作用 ..... | 9   |
| 1.5.2 microRNA 在角膜的表达及功能研究进展.....  | 9   |
| 1.5.3 microRNA 在皮肤的表达及功能研究进展.....  | 11  |
| 1.6 p38MAPK 与 p63.....             | 12  |
| 1.6.1 p38MAPK 信号通路 .....           | 12  |
| 1.6.2 p63.....                     | 13  |
| 1.7 课题背景意义 .....                   | 14  |
| 第二章 实验材料和方法 .....                  | 16  |
| 2.1 实验药品、试剂与仪器 .....               | 16  |
| 2.1.1 人体组织标本.....                  | 16  |
| 2.1.2 细胞株.....                     | 16  |
| 2.1.3 主要试剂和材料.....                 | 16  |
| 2.1.4 主要实验仪器和耗材.....               | 17  |

|   |           |
|---|-----------|
| <b>2.2 实验方法 .....</b>                                 | <b>18</b> |
| 2.2.1 细胞学相关实验.....                                    | 18        |
| 2.2.2 分子生物及组织化学相关实验.....                              | 22        |
| 2.2.3 microRNA 靶基因的预测.....                            | 33        |
| 2.2.4 统计学分析.....                                      | 34        |
| <b>第三章 实验结果和讨论 .....</b>                              | <b>35</b> |
| <b>3.1 实验结果 .....</b>                                 | <b>35</b> |
| 3.1.1 正常人角膜缘干细胞和皮肤干细胞 microRNA 的差异性表达.....            | 35        |
| 3.1.2 荧光实时定量 PCR 验证基因芯片结果.....                        | 37        |
| 3.1.3 角膜和皮肤上皮干细胞分化过程中差异性表达的 microRNA 的变化.....         | 40        |
| 3.1.4 miR-145 在人角膜缘和皮肤上皮的基底细胞高表达 .....                | 43        |
| 3.1.5 miR-145 抑制人角膜和皮肤上皮细胞增殖 .....                    | 44        |
| 3.1.6 miR-145 可能通过抑制 p38 的表达, 使其信号通路下游的 p63 表达下降..... | 46        |
| <b>3.2 讨论 .....</b>                                   | <b>47</b> |
| 3.2.1 角膜缘上皮干细胞和皮肤上皮干细胞中差异性表达的 microRNA .....          | 49        |
| 3.2.2 角膜上皮干细胞和皮肤上皮干细胞分化过程中差异性 microRNA 的变化及意义探讨.....  | 51        |
| 3.2.3 miRNA-145 在角膜和皮肤上皮干细胞中功能和作用机制探讨.....            | 52        |
| <b>第四章 结论 .....</b>                                   | <b>55</b> |
| <b>问题与展望 .....</b>                                    | <b>56</b> |
| <b>参 考 文 献 .....</b>                                  | <b>57</b> |
| <b>致 谢.....</b>                                       | <b>63</b> |



## Catalogue

|  |            |
|--|------------|
| <b>Abbreviation.....</b>   | <b>V</b>   |
| <b>Abstract.....</b>   | <b>VII</b> |
| <b>Chapter 1 Introduction.....</b>   | <b>1</b>   |
| <b>1.1 Background Information of microRNAs .....</b>   | <b>1</b>   |
| 1.1.1 Discovery and Function of microRNAs .....  | 1          |
| <b>1.2 Anatomy and Physiology of Normal Cornea .....</b>   | <b>3</b>   |
| <b>1.3 Corneal Epithelial Cells and Skin Epithelial Cells .....</b>                                | <b>4</b>   |
| 1.3.1 Similarities and Differences between Corneal Epithelial Cells and Skin Epithelial Cells..... | 4          |
| 1.3.2 Mutual Transformation between Corneal Epithelial Cells and Skin Epithelial Cells.....        | 5          |
| <b>1.4 Limbal Epithelial Stem Cells and Skin Epithelial Stem Cells.....</b>                        | <b>6</b>   |
| 1.4.1 Limbal Epithelial Stem Cells .....   | 6          |
| 1.4.2 Skin Epithelial Stem Cells .....   | 8          |
| <b>1.5 Research Progress of microRNAs in Cornea and Skin .....</b>                                 | <b>9</b>   |
| 1.5.1 Vital Roles of microRNAs in Cornea and Skin .....  | 9          |
| 1.5.2 Research Progress of microRNA Expression and Function in Cornea...                           | 9          |
| 1.5.3 Research Progress of microRNA Expression and Function in Skin .....                          | 11         |
| <b>1.6 p38 MAPK and p63 .....</b>  | <b>12</b>  |
| 1.6.1 p38 MAPK Signaling Pathway .....   | 12         |
| 1.6.2 p63.....   | 13         |
| <b>1.7 Background Significance of this Project .....</b>   | <b>14</b>  |
| <b>Chapter 2 Materials and Methods .....</b>   | <b>16</b>  |
| <b>2.1 Reagents and Instruments.....</b>   | <b>16</b>  |
| 2.1.1 Human Tissue Samples .....   | 16         |
| 2.1.2 Cell Lines .....   | 16         |
| 2.1.3 Reagents and Materials .....   | 16         |

|   |           |
|---|-----------|
| 2.1.4 Instruments and Expendable Supplies .....   | 17        |
| <b>2.2 Protocols and Recipes .....</b>  | <b>18</b> |
| 2.2.1 Cytological Experimental Methods .....  | 18        |
| 2.2.2 Molecular Biological Technologies .....   | 22        |
| 2.2.3 Prediction of microRNA Target Genes .....   | 33        |
| 2.2.4 Statistical Analysis .....  | 34        |
| <b>Chapter 3 Results and Discussions.....</b>   | <b>35</b> |
| <b>3.1 Results .....</b>  | <b>5</b>  |
| 3.1.1 miRNA arrays identification of differentially expressed microRNAs<br>between LESC and SESC .....          | 35        |
| 3.1.2 real-time PCR verification of miRNA arrays results .....  | 37        |
| 3.1.3 microRNAs expression changes during LESC and SESC<br>differentiation.....                                 | 40        |
| 3.1.4 miR-145 expresses in the basal cells of limbus and skin.....  | 43        |
| 3.1.5 miR-145 inhibits proliferation of skin and corneal epithelial cells.....                                  | 44        |
| 3.1.6 miR-145 might target p38 to regulate p63 expression.....  | 46        |
| <b>3.2 Discussion.....</b>  | <b>47</b> |
| 3.2.1 Significance of differentially expressed microRNAs between LESC<br>and SESC.....                          | 49        |
| 3.2.2 Significance of the identified microRNAs expression changes during<br>LESC and SESC differentiation ..... | 51        |
| 3.2.3 Functions and possible mechanism of miR-145 in LESC and SESC  | 52        |
| <b>Chapter 4 Conclusion .....</b>   | <b>55</b> |
| <b>Questions and Prospects.....</b>   | <b>56</b> |
| <b>References .....</b>   | <b>57</b> |
| <b>Acknowledgement.....</b>   | <b>63</b> |

英文缩略语对照表

| Abbreviation          | Full Name  |
|-----------------------|--|
| BSA                   | albuminbovine  |
| cDNA                  | complementary DNA  |
| DMSO                  | dimethyl sulfoxide   |
| EDTA                  | ethylene diamine tetraacetic acid                            |
| EGF                   | epidermal growth factor                                      |
| FAM                   | carboxyfluorescein   |
| FBS                   | fetal bovine serum   |
| HCE                   | human corneal epithelial cells                               |
| HEPES                 | N-(2-hydroxyethyl)piperazine-N-2-ethancsulfonic acid         |
| ITS                   | insulin-transferrin-selenium                                 |
| K1/3/5/10/12/14/15/19 | cytokeratin 1 /3/5/10/12/14/15/19                            |
| KSFM                  | keratinocyte serum-free medium                               |
| LESCs                 | limbal epithelial stem cells                                 |
| MMC                   | mitomycin C  |
| MTT                   | 3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide |
| microRNA              | micro ribonucleic acid                                       |
| mRNA                  | messsage ribonucleic acid                                    |
| NC                    | negative control   |
| Pax6                  | paired box gene 6  |
| PBS                   | phosphate buffered saline                                    |
| PFA                   | paraformaldehyde   |
| PVDF                  | immobilon-P transfer membrane                                |
| p38                   | p38 mitogen-activated protein kinases                        |
| p63                   | tumor protein 63   |
| Realtime PCR          | realtime polymerase chain reaction                           |

英文缩略语对照表

|        |   |
|--------|---|
| RT-PCR | reverse transcription polymerase chain reaction |
| SDS    | sodium dodecyl sulfate                          |
| SESCs  | skin epithelial stem cells                      |
| SHEM   | supplemental hormonal epithelial media          |
| SSC    | sodium chloride-sodium citrate                  |
| TEMED  | tetramethylethylenediamine                      |
| UTR    | untranslated region                             |

## 摘 要

**目的:**通过基因芯片技术找出角膜上皮干细胞和皮肤上皮干细胞差异性表达的 microRNAs 并对其在增殖和分化方面的功能及作用机制进行初步的研究和探讨。

**方法:**

1. 运用克隆培养方法分别培养三组人角膜和皮肤上皮干细胞, 进行 microRNA 基因芯片分析, 找出有显著差异性表达的 microRNA ( $p \leq 0.05$  差异倍数  $\geq 1.5$ )。
2. 对基因芯片分析出的差异性表达的 microRNA 进行荧光实时定量 PCR, 筛选出表达模式与芯片结果一致并且差异倍数较大的 microRNA。
3. 运用荧光实时定量 PCR, 研究筛选出来的有显著差异的 microRNA 在角膜和皮肤上皮干细胞分化过程中的变化。
4. 利用瞬时转染的方法将在角膜缘干细胞中显著表达上调的 miR-145 mimics 转染进入皮肤上皮干细胞, 用相差显微镜观察细胞形态变化, MTT 检测细胞增殖能力的变化。
5. 用 TargetScan 及 miRanda 预测 miR-145 与增殖相关的靶基因, 荧光实时定量 PCR 和蛋白免疫印迹 (Western blot) 研究证实过表达 miR-145 后其靶基因及靶基因下游基因的变化。

**结果:**

1. microRNA 基因芯片分析共找出了 27 个在角膜上皮干细胞和皮肤上皮干细胞中差异性表达的 microRNA; 其中在角膜上皮干细胞中表达较高的有 17 个, 为 miR-30a, miR-1297, miR-4301, miR-4262, miR-199aa-3p, miR-190, miR-302f, miR-338-5p, miR-484, miR-21, miR-376b, miR-26a, miR-140-3p, miR-18b, miR-145, miR-652, miR-324-5p; 在皮肤上皮干细胞中表达较高的 10 个, 为 miR-23a\*, miR-943, miR-1236, miR-193b\*, miR-382, miR-542-3p, miR-205\*, miR-224\*, miR-378, miRPlus-1382\*。
2. 实时荧光定量 PCR 验证出与基因芯片结果表达模式一致且差异倍数较大的 microRNA 有 8 个 (miR-4262, miR-199aa-3p, miR-190, miR-302f, miR-21, miR-376b, miR-140-3p, miR-145), 均为在角膜缘上皮干细胞中显著上调表达的 microRNA。
3. 对角膜和皮肤干细胞分化过程筛选出的 8 个差异性表达的 microRNA 变化的

分析中发现,其中 6 个在角膜和皮肤上皮干细胞分化过程中表达均显著降低,而仅 miR-302f 和 miR-4262 在角膜上皮干细胞分化过程中显著升高,而在皮肤上皮干细胞分化过程中显著降低。

4. 选取 miR-145 进行初步的功能研究,结果表明 miR-145 可以抑制皮肤上皮干细胞的增殖能力。
5. miR-145 抑制皮肤上皮干细胞增殖能力的功能是通过下调 p38 基因的表达从而抑制 p38 MAPK 信号通路来实现的,同时信号通路下游基因 p63 的表达也降低。

#### 结论:

1. 差异性表达的 microRNA 在维持角膜和皮肤上皮干细胞各自的特性中可能起到关键作用。
2. 差异性表达的 microRNA 在角膜和皮肤上皮干细胞分化过程中的变化提示其在维持上皮干细胞干性和引导不同上皮干细胞特定分化过程中的重要作用。
3. miR-145 调控皮肤及角膜上皮细胞的增殖能力,这在对上皮细胞的异常增殖和成瘤的控制中可能起着重要作用。

**关键词:** 角膜; 皮肤; microRNA

## Abstract

**Objective** To identify the differentially expressed microRNAs between limbal epithelial stem cells (LESCs) and skin epithelial stem cells (SESCs) and investigate the functions of one of these microRNAs.

### Methods

1. 6 samples of clone cultured limbal epithelial stem cell and skin epithelial stem cell (each 3 samples respectively) were collected and analyzed by microRNA array. To identify differentially expressed miRNAs with statistical significance, we performed a volcano plot filtering between the two groups from the experiment. The threshold we used to screen up or down regulated miRNAs is fold change  $\geq 1.5$  and P value  $\leq 0.05$ .
2. Verify the miRNA arrays identified differentially expressed miRNAs by realtime PCR, and select those miRNAs with the same expression pattern and prominent fold change.
3. Investigate the change of the selected miRNAs during differentiation of LESCs and SESC by real-time qPCR.
4. Transfect one of the selected miRNAs---miR-145 into SESC, observing morphologic changes by phase contrast microscope and detecting proliferating ability by MTT.
5. Target genes of miR-145 related to cell proliferation were predicted by TargetScan and miRanda. Expression changes of the predicted gene and the downstream gene after miR-145 transfection were detected by realtime PCR and Western blot.

### Results

1. 27 differentially expressed microRNAs were identified by miRNA array; 17 miRNAs were up regulated in LESCs, which are miR-30a, miR-1297, miR-4301, miR-4262, miR-199a-3p, miR-190, miR-302f, miR-338-5p, miR-484, miR-21, miR-376b, miR-26a, miR-140-3p, miR-18b, miR-145,

miR-652, miR-324-5p; 10 miRNAs were up regulated in SESC, which are miR-23a\*, miR-943, miR-1236, miR-193b\*, miR-382, miR-542-3p, miR-205\*, miR-224\*, miR-378, miRPlus-1382\*.

2. 8 miRNAs (miR-4262, miR-199a-3p, miR-190, miR-302f, miR-21, miR-376b, miR-140-3p, miR-145) were selected by realtime PCR with the same expression pattern and prominent fold change, all of which are up regulated in LESC.
3. 6 of the 8 selected differentially expressed miRNAs were down regulated remarkably during both LESC and SESC differentiation, while 2 miRNAs, miR-302f and miR-4262 were up regulated dramatically during LESC differentiation but down regulated dramatically during SESC differentiation.
4. miR-145 was selected to undergo function investigation and results showed proliferative inhibition by miR-145 over-expression in SESC.
5. Further mechanism studies indicated that the cell proliferative inhibition of miR-145 is through inhibition of p38 MAPK signaling pathway by down regulating miR-145 target gene p38, and along with down regulating of p38 MAPK downstream gene p63.

## Conclusion

1. The differentially expressed miRNAs between LESC and SESC might play vital roles in keeping their stemness and respective properties.
2. The different changes of the selected differentially expressed miRNAs during LESC and SESC differentiation indicated their functions in directing respective differentiations of LESC and SESC.
3. Proliferative inhibition of miR-145 in corneal and skin epithelial cell indicated its role in control of proliferation and tumorigenesis.

**Keywords** Limbal epithelial stem cells; Skin epithelial stem cells; microRNAs



Degree papers are in the "[Xiamen University Electronic Theses and Dissertations Database](#)". Full texts are available in the following ways:

1. If your library is a CALIS member libraries, please log on <http://etd.calis.edu.cn/> and submit requests online, or consult the interlibrary loan department in your library.
2. For users of non-CALIS member libraries, please mail to [etd@xmu.edu.cn](mailto:etd@xmu.edu.cn) for delivery details.

厦门大学博硕士论文摘要库